# Endotoxin Assay Kit Instruction for use









REF: THG10-0030, THG10-0060, THG10-0125, THG10-0250, THG10-0500

Product Name: Endotoxin Assay Kit, Gel-Clot

Specifications: 44 tests/box

# Intended use:

- For the qualitative detection of endotoxin with gel-clot method on pharmaceuticals, biological products, radioactive medicine, and medical device.
- For the pyrogen control or endotoxin detection during the production.
- Other situation needs detection of endotoxin with gel-clot method.

## Principle:

The clotting protein and clotting enzyme in the TAL (Tachypleus Amebocyte Lysate) could be activated by very small dose of endotoxin. Then TAL and bacterial endotoxin could agglutinate and come into gel. So it is an agglutination to detect the endotoxin by qualitation.

# Materials supplied with the Endotoxin Assay kit:

Item No.	Product Name	Description					
THG10-0030	Endotoxin Assay Kit	TAL(0.03EU/mL) 0.2mL×44pcs; BET water 2mL×5pcs					
	(Gel-Clot)	Control Standard Endotoxin ×1pcs;					
	Endotoxin Assay Kit	TAL(0.06EU/mL) 0.2mL×44pcs;					
THG10-0060	(Gel-Clot)	BET water 2mL×5pcs					
	(der-Ciot)	Control Standard Endotoxin ×1pcs;					
THG10-0125	F-1-4 V:4	TAL(0.125EU/mL) 0.2mL×44pcs;					
	Endotoxin Assay Kit	BET water 2mL×5pcs					
	(Gel-Clot)	Control Standard Endotoxin ×1pcs;					
	E-1-4i- A V:4	TAL(0.25EU/mL) 0.2mL×44pcs;					
THG10-0250	Endotoxin Assay Kit	BET water 2mL×5pcs					
	(Gel-Clot)	Control Standard Endotoxin ×1pcs;					
THG10-0500	Endotovin Accountit	TAL(0.50EU/mL) 0.2mL×44pcs;					
	Endotoxin Assay Kit	BET water 2mL×5pcs					
	(Gel-Clot)	Control Standard Endotoxin ×1pcs;					

# Materials required but not supplied

Vortex mixer; Laboratory Water Bath (incubator, the temperature could be set to  $37\pm1^{\circ}$ ); Test tube rack, Adjustable Pipette, Laboratory Film (Parafilm), 75% alcohol swab, endotoxin-free tube and endotoxin-free pipette tips.

Note: endotoxin-free tube and endotoxin-free pipette tips could be purchased separately.

Page 1

## Reagent Storage

Store all reagents, as supplied, at 0-25°C in cool condition without corrosive gas and away from light, the related humidity should be less than 80%. The expiry date is 24 months. Use immediately after opening.

### Procedures:

- 1. Check the sensitivity of TAL if necessary
- 1.1 Preheat the incubator to  $37 \pm 1^{\circ}$ C
- 1.2 Preparation of the control standard endotoxin solution:

Take out the Control standard endotoxin, clean the bottle neck with 75% alcohol and open it by breaking apart it from the bottle neck, add 1mL BET water, mix for 15min with Vortex mixer, as per the sensitivity( $\lambda$ ) of the TAL dilute it to 2  $\lambda$  ,  $\lambda$  , 0.5  $\lambda$  and 0.25  $\lambda$  , mix with Vortex mixer for 30s for each step. I.e.  $\lambda$  =0.125EU/mL, then dilute the control standard endotoxin to 0.25EU/mL, 0.125EU/mL, 0.0625EU/mL and 0.0312EU/mL.

- \* After reconstitution before any dilution, the CSE can be sealed and stored in refrigerator at -10C for 15days. When you need to use it, take it out and unfreeze it at room temperature and then Vortex for 15min.
- 1.3 Preparation of the TAL solution and application of the samples.

Take out 18pcs TAL, put them on the test tube rack, and application samples according to below table:

Sensitivity of the endotoxin (EU/ml)₽	0.25∉			0.125₽			0.0625₽			0.0312∉			Negative control₽					
TAL₽	⊕@	@₽	3₽	@₽	⊕	@₽	3₽	@.₽	⊕	20	3₽	@.	⊕	Q.	3₽	@.₽	⊕	②₽
Endotoxin Detection water(ml)	-4	-0	-4	-4	-0	=Ø3	- <i>Q</i>	-0	-4	-42	-0	-47	-0	-0	-φ	-47	0.2₽	0.2₽
Control standard endotoxin(ml)	0.2₽	0.2₽	0.20	0.2₽	0.2₽	0.2₽	0.2₽	0.2₽	0.2₽	0.2₽	0.2₽	0.20	0.20	0.2₽	0.2₽	0.2	-0	-0

After application the samples, mix them even with Vortex mixer, cover the tube with laboratory film(Parafilm), and put them into incubator which setting the temperature at  $37\pm1^{\circ}\mathrm{C}$ . Last for  $60\mathrm{min}\pm2\mathrm{mins}$ . Then carefully remove the tubes one by one from the incubator. Gently invert the tubes to 180 degree, if a solid clot has formed the result is positive (+), if no clot has formed, the result is negative (-). Do not shake the tubes when reading the test.

# The test is valid only when:

Both negative tubes: no clot has formed (-)

0.25λ tubes: no clot has formed (-)

2λ tubes: Should clot (+)

Calculate the sensitivity of the TAL( $\lambda c$ ) via below formula:

 $\lambda c = Ig^{-1}(\sum X/4)$ 

X = lg (highest sensitivity appeared to be positive (+))

Only when the value of  $\lambda c$  is between 0.5 $\lambda$  to 2 $\lambda$ , the TAL could be used for testing, and please use the nominated sensitivity. I.e. according to the test result in below table,

	0.25(2λ)	0.125(λ)	0.0625(0.5λ)	0.0312(0.25λ)	Negative	highest sensitivity		
					control	appeared to be (+)		
1	+	+	-	-	-	0.125		
2	+	+	-	-	-	0.125		
3	+	+	-	-		0.125		
4	+	+	-	-		0.125		

 $\lambda c = Ig^{-1}(\sum X/4)$ 

 $= \lg^{-1} ((\lg 0.125 + \lg 0.125 + \lg 0.125 + \lg 0.125)/4)$ 

=0.125EU/mL

- 2. Sample testing
- 2.1 Preheat the incubator to 37±1°C
- 2.2 Preparation of the positive control solution:

Take out the Control standard endotoxin, clean the bottle neck with 75% alcohol and open it by breaking apart it from the bottle neck, add 1mL BET water, mix for 15min with Vortex mixer and dilute it to  $2\lambda$  and  $4\lambda$  ( $\lambda$  is the sensitivity of the TAL).

Example: If the sensitivity of the Control standard endotoxin is 10EU/mL, and  $\lambda=0.125EU/mL$  then please follow the below steps:

$$10EU/ml \xrightarrow{0.2ml} 1.0EU/ml \xrightarrow{1.0ml} 1.0ml > 0.50EU/ml \xrightarrow{0.5ml} 0.25EU/ml$$

Above volume is the amount being taken out, while the below volume is the amount we add BET water.

2.3 Preparation of the sample:

Calculate Maximum valid dilution (MVD)= cL/λ

- L: Endotoxin limit
- c: Density of the sample, when the unit of L is EU/mL, then c=1.0mL/mL; when the unit of L is EU/mg or EU/U, then the unit of c should be mg/mL or U/mL;
- λ: Sensitivity (EU/mL) of TAL

Example: If endotoxin limit is 0.5EU/mL, the sample is glucose injection,  $\lambda$ =0.125EU/mL then MVD=1.0\*0.50/0.125=4

Then take out 0.50mL sample and put into 1.50mL BET water, mix even.

Note: The pH value of the specimen should be within 6-8, if not, the pH buffer should be used. Always measure the pH of the bulk sample to avoid contamination by the pH electrode

2.4 Preparation of the positive product control solution:

Mix the 0.5mL Control standard endotoxin solution ( $4\lambda$ ) and 0.5mL 2 times diluted sample solution (if it is glucose injection, take out 0.25mL and add 0.25mL BET water, mix even) with Vortex mixer.

2.5 Preparation of the reagent:

According to the quantity of the samples, take out the TAL, clean the bottle with 75% alcohol and open it by breaking apart it from the bottle neck. Put them on the test tube rack.

2.6 Application of samples: Refer to the table below for application of samples, and mix even by Vortex.

MOX.										
	Sample tube		Negative	e control	Positive	product	Positive control			
	1 2		1 2		1)	2	1)	2		
TAL	1pcs	1pcs	1pcs	1pcs	1pcs	1pcs	1pcs	1pcs		
BET water			0.2	0.2						
Samples	0.2	0.2								
Control										
standard							0.2	0.2		
endotoxin							0.2	0.2		
solution(2λ)										
Positive										
product					0.2	0.2				
control					0.2	0.2				
solution										

- 2.7 Cover the tubes with Laboratory Film, then put it into the 37°C±1 incubator for 60±2mins.
- 2.8 Then carefully remove the tubes one by one from the incubator. Gently invert the tubes to 180

degree, if a solid clot has formed the result is positive (+), if no clot has formed, the result is negative (-). Do not shake the tubes when reading the test.

2.9 Interpretation of the result:

The test is valid only when:

Both negative tubes: no clot has formed (-)

Both Positive product control tubes: should clot (+)

Both positive control tubes: Should clot (+)

The sample tubes:

If both negative (-) means the sample contains an endotoxin concentration less than  $\lambda^*MVD$ ; If both positive (+), means the sample contains an endotoxin concentration of at least  $\lambda^*MVD$ . If one is negative (-), and the other is positive (+), then retest the sample. And 4 sample tubes needed, all 4 are negative means qualified, else not qualified.

## Precaution

- The pH value of the sample solution should be within 6-8, if not, the pH buffer should be used. Always
  measure the pH of the bulk sample to avoid contamination by the PH electrode. pH buffer could be
  purchased separately.
- 2. All materials used for specimen collection and test reagent preparation should be endotoxin-free.
- 3. Mix always with the Vortex mixer.
- 4. Do not shake the tubes when reading the test.
- 5. If interference test is necessary, please refer to National Pharmacopeia.
- 6. Specimens should be clearly labeled during the test.
- 7. Main reagent is made from lyophilized TAL (Tachypleus Amebocyte Lysate); which occasionally causes some people sneezing and/or watery eyes symptoms of allergy after contact. Please stop the operation and move to ventilation and open place if such uncomfortable symptoms occur.

# Reference

- 1. Kun Wang, The application value of detect of bacterial endotoxin in patients with infectious disease in ICU ward. China Modern Medicine. 2014, 01(b)-0125-02.
- Van Deventer SJ, Buller HR, ten Cate JW, et al. Endotoxaemia: an early predictor of septicaemia in febrile patients[J]. Lancet, 1988, 1(9596):605-609.
- 3. Ling Wang, Guiping Wang. Endotoxin detecting method and the application summary[J]. China Pharmacist, 2003, 6(5): 316-317.
- 4. Annex II E Bacterial Endotoxins Test (Kinetic-Turbidimetric method). 2010



Zhanjiang Bokang Marine Biological co., ltd.

#188, RuiYunNan Rd., MaZhang district, Zhan Jiang, Guang Dong, China

Page 4

Zipcode: 524094

Page 3